Fenics **BIO**

Product data sheet

Raw264.7/GFP stable cell line Catalog Number: CL-1133 Storage: Liquid nitrogen Components: 1 vial contains ~2 x10⁶ cells in Cell freezing medium

Product description

Raw264.7/GFP cells are derived from the mouse macrophage cell line Raw264.7 by stably integration of a constitutive GFP expression construct. Raw264.7 cells have been used as a model for studying the response of inflammatory molecules to various stimuli. Raw264.7/GFP cells stably express GFP, can be used for *in vitro* assays and *in vivo* imaging.

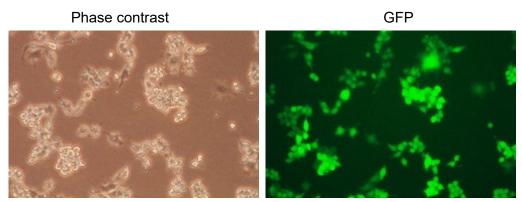


Figure 1. GFP expression in Raw264.7/GFP stable cell line

Cell line description

Organism: *Mus musculus,* mouse Tissue: Ascites Cell type: macrophage Morphology: monocyte/macrophage Culture Properties: adherent Disease: Abelson murine leukemia virus-induced tumor Biosafety Level: 2

Medium

- Complete culture medium: DMEM with 10% fetal bovine serum (FBS)
 1 μg/mL of puromycin may be added to the culture medium. Puromycin should not be added until a culture has been well established from the thawed cells.
- 2. Freeze medium: FBS with 6% DMSO

Culture procedure

Thawing of frozen cells

- 1. Thaw the frozen cryovial by gentle agitation in a 37 °C water bath in 1-2 minutes.
- 2. Remove the cryovial from the water bath as soon as the contents are thawed and decontaminate by wiping with 70% ethanol.
- 3. Transfer the thawed cell suspension to a centrifuge tube containing 10 ml of Complete culture medium, centrifuge at 500 g for 5 minutes.
- 4. Remove the medium by aspiration, resuspend the cells with 10 ml of the Complete culture medium by gently pipetting up and down.
- 5. Transfer the cells to a 10 cm cell culture dish.
- 6. Place the cells in a 37°C incubator with 5% CO2.

Sub-culturing

Volumes are given for a 10 cm cell culture dish. Increase or decrease the amount of dissociation medium needed proportionally.

- 1. Remove the medium by aspiration.
- 2. Briefly rinse the cell layer with 1xDPBS to remove all traces of serum that contains trypsin inhibitor.
- 3. Add 1 ml of Trypsin-EDTA (0.25%) solution to the dish and observe cells under an inverted microscope until cell layer is dispersed.
- 4. Add 4 ml of complete growth medium and aspirate cells by gently pipetting.
- 5. Add appropriate aliquots of the cell suspension to new culture vessels. Incubate cultures at 37°C with 5% CO2.