# Fenics **BIO**

### Product data sheet

HepG2/GFP stable cell line Catalog Number: CL-1109 Storage: Liquid nitrogen Components: 1 vial contains ~2 x10<sup>6</sup> cells in Cell freezing medium

## **Product description**

HepG2/GFP cells are derived from the human hepatocarcinoma HepG2 cell line by stably integration of a constitutive GFP stably expression construct. The HepG2 cell line has been widely used as a model system in biomedical research, particularly in studies related to liver biology, drug metabolism, toxicology, hepatitis virus infection and liver cancer. HepG2/GFP cells stably express GFP, can be used for in vitro assays and in vivo imaging.

Phase contrast

GFP

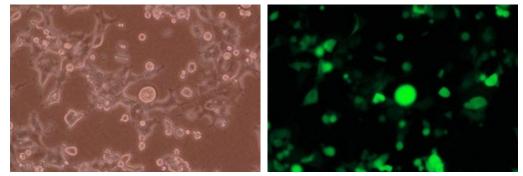


Figure 1. GFP expression in HepG2/GFP stable cell line

#### **Cell line description**

Organism: Homo sapiens (human) Tissue: Liver Morphology: epithelial-like Culture Properties: adherent Disease: Carcinoma; Hepatocellular Biosafety Level: 2

#### Medium

1. Complete culture medium: EMEM with 10% fetal bovine serum (FBS)

 $2~\mu g/mL$  of puromycin may be added to the culture medium. Puromycin should not be added until a culture has been well established from the thawed cells.

2. Freeze medium: FBS with 6% DMSO

#### Culture procedure

#### Thawing of frozen cells

- 1. Thaw the frozen cryovial by gentle agitation in a 37 °C water bath in 1-2 minutes.
- 2. Remove the cryovial from the water bath as soon as the contents are thawed, and decontaminate by wiping with 70% ethanol.
- 3. Transfer the thawed cell suspension to a centrifuge tube containing 10 ml of Complete culture medium, centrifuge at 500 g for 5 minutes.
- 4. Remove the medium by aspiration, resuspend the cells with 10 ml of the Complete culture medium by gently pipetting up and down.
- 5. Transfer the cells to a 10 cm cell culture dish.
- 6. Place the cells in a 37°C incubator with 5% CO2.

#### Sub-culturing

Volumes are given for a 10 cm cell culture dish. Increase or decrease the amount of dissociation medium needed proportionally.

- 1. Remove the medium by aspiration.
- 2. Briefly rinse the cell layer with 1xDPBS to remove all traces of serum that contains trypsin inhibitor.
- 3. Add 1 ml of Trypsin-EDTA (0.25%) solution to the dish and observe cells under an inverted microscope until cell layer is dispersed.
- 4. Add 4 ml of complete growth medium and aspirate cells by gently pipetting.
- 5. Add appropriate aliquots of the cell suspension to new culture vessels. Incubate cultures at 37°C with 5% CO2.

FenicsBIO 1448 South Rolling Rd. Halethorpe, MD 21227, USA www.FenicsBIO.com info@FenicsBIO.com Tel: 1-800-811-8291